AlphaLISA® Research Reagents

Bovine Immunoglobulin G2 (bIgG2) AlphaLISA Immunoassay Kit

Product number: AL532 HV/C/F

Research Use Only. Not for use in diagnostic procedures.

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Product Information

Application: This kit is designed for the quantitative determination of bovine IgG2 in buffer using a

homogeneous AlphaLISA assay (no wash steps). The assay shows negligible cross-reactivity

with other bovine Igs and IgGs from othe sepcies.

Sensitivity: Lower Detection Limit (LDL): 2.06 ± 1.3 ng/mL

Lower Limit of Quantification (LLOQ): 5.67 ng/mL

EC₅₀: 179 ± 60 ng/mL

Dynamic range: 2.1 - 3000 ng/mL (Figure 1).

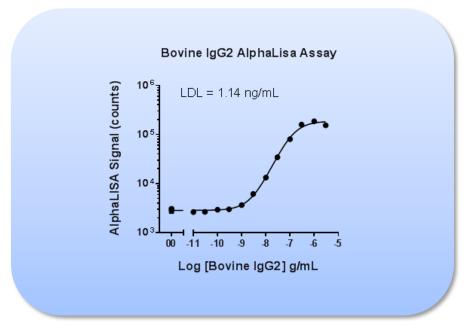


Figure 1. Typical sensitivity curves in AlphaLISA Bovine Immunoassay Buffer. The data was generated using a white OptiplateTM-384 microplate and the EnVision[®] Multilabel Plate Reader with Alpha option 2102.

Storage: Store kit in the dark at +4°C. Store reconstituted analyte at -20°C.

Stability: This kit is stable for at least 6 months from the manufacturing date when stored in its original

packaging and the recommended storage conditions. Note: Once reconstituted, the bovine IgG2

analyte is stable for at least 18 months when stored at -20°C.

Quality Control

Lot to lot consistency is confirmed in an AlphaLISA assay. Maximum and minimum signals, EC₅₀ and LDL were measured on the EnVision Multilabel Plate Reader with Alpha option using the protocol described in this technical data sheet. We certify that these results meet our quality release criteria. Maximum counts may vary between bead lots and the instrument used, with no impact on LDL measurement.

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Analyte of Interest

There are five classes of mammalian immunoglobulins: IgA, IgD, IgE, IgM, and IgG. IgG is the most abundant immunoglobulin and is equally distributed in blood and tissue. In bovine, the IgG class is further divided into two subclasses: IgG1 and IgG2. The general immunoglobulin structure is composed of four polypeptide chains, two heavy and two light chains linked together and to each other by disulfide bonds, creating a tetrameric quaternary structure. IgG1 is involved in response to a foreign antigen. The presence of IgG2 usually signifies a mature antibody response. IgG2 has a molecular weight of about 150 kDa, it can bind to many pathogens and also plays an important role in antibody-dependent cell-mediated cytotoxicity. Typically, bovine serum and plasma samples contain about 5.0 to 13.5 mg/ml of IgG2.

Description of the AlphaLISA Assay

AlphaLISA technology allows the detection of molecules of interest in buffer, cell culture media, serum and plasma in a highly sensitive, quantitative, reproducible and user-friendly mode. In an AlphaLISA assay, a Biotinylated Anti-Analyte Antibody binds to the Streptavidin-coated Alpha Donor beads, while another Anti-Analyte Antibody is conjugated to AlphaLISA Acceptor beads. In the presence of the analyte, the beads come into close proximity. The excitation of the Donor beads provokes the release of singlet oxygen molecules that triggers a cascade of energy transfer in the Acceptor beads, resulting in a sharp peak of light emission at 615 nm (Figure 2).

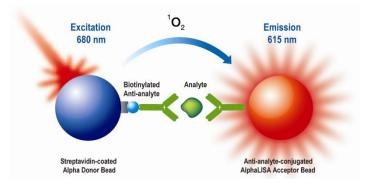


Figure 2. AlphaLISA Assay principle.

Precautions

- The AlphaScreen® Donor beads are light-sensitive. All the other assay reagents can be used under normal light conditions. All Alpha assays using the Donor beads should be performed under subdued laboratory lighting (< 100 lux). Green filters (LEE 090 filters (preferred) or Roscolux filters #389 from Rosco) can be applied to light fixtures.
- All blood components and biological materials should be handled as potentially hazardous. The analyte included in this kit is from a bovine source.
- Some analytes are present in saliva. Take precautionary measures to avoid contamination of the reagent solutions.
- The Biotinylated Anti-Analyte Antibody contains sodium azide. Contact with skin or inhalation should be avoided.

Kit Content: Reagents and Materials

Kit components	AL532HV (100 assay points***)	AL532C (500 assay points***)	AL532F (5 000 assay points***)
AlphaLISA Anti-Bovine IgG2 Acceptor beads stored in PBS, 0.05% Proclin- 300, pH 7.2	20 μL @ 5 mg/mL (1 brown tube, <u>white</u> cap)	50 μL @ 5 mg/mL (1 brown tube, <u>white</u> cap)	500 μL @ 5 mg/mL (1 brown tube, <u>white</u> cap)
Streptavidin (SA)-coated Donor beads stored in 25 mM HEPES, 100 mM NaCl, 0.05% Proclin-300, pH 7.4	100 μL @ 5 mg/mL (1 brown tube, black cap) 200 μL @ 5 mg/mL (1 brown tube, black cap)		2 mL @ 5 mg/mL (2 brown tubes, <u>black</u> caps)
Biotinylated Antibody Anti-Bovine IgG2 stored in PBS, 0.1% Tween-20, 0.05% NaN ₃ , pH 7.4	50 μL @ 500 nM (1 tube, <u>black</u> cap)	120 μL @ 500 nM (1 tube, <u>black</u> cap)	1 mL @ 500 nM (1 tube, <u>black</u> cap)
AlphaLISA Bovine IgG2 (3 μg), lyophilized analyte *	1 tube, <u>clear</u> cap	1 tube, <u>clear</u> cap	1 tube, <u>clear</u> cap
AlphaLISA Bovine Immunoassay Buffer (10X)	2.5 mL, 1 small bottle	10 mL, 1 small bottle	100 mL, 1 large bottle

^{*} Reconstitute bovine IgG2 in 100 μL Milli-Q® grade H₂O. The reconstituted analyte should be used within 60 minutes or aliquoted into screw-capped polypropylene vials and stored at -20°C for further experiments. Avoid multiple freeze-thaw cycles. It has been demonstrated that reconstituted bovine IgG2 is stable for at least 18 months at -20°C. One vial contains an amount of bovine IgG2 sufficient for performing 10 standard curves. Additional vials can be ordered separately (cat # AL532S).

Sodium azide should **not** be added to the stock reagents. High concentrations of sodium azide (> 0.001 % final in the assay) might decrease the AlphaLISA signal. Note that sodium azide from the Biotinylated Antibody stock solution will not interfere with the AlphaLISA signal (0.0001% final in the assay).

Specific additional required reagents and materials:

The following materials are recommended:

Item	Suggested source	Catalog #
TopSeal™-A Adhesive Sealing Film	PerkinElmer Inc.	6050195
EnVision®-Alpha Reader	PerkinElmer Inc.	-



^{***} The number of assay points is based on an assay volume of 100 μL in 96-well plates (AL532HV) or 50 μL in 96- or 384-well assay plates using the kit components at the recommended concentrations.

Recommendations

General recommendations:

- The volume indicated on each tube is guaranteed for single pipetting. Multiple pipetting of the reagents may reduce the theoretical amount left in the tube. To minimize loss when pipetting beads, it is preferable not to pre-wet the tip.
- Centrifuge all tubes (including lyophilized analyte) before use to improve recovery of content (2000g, 10-15 sec).
 Re-suspend all reagents by vortexing before use.
- Use Milli-Q® grade H_2O (18 $M\Omega$ •cm) to dilute 10X AlphaLISA Bovine Immunoassay Buffer to reconstitute the lyophilized analyte.
- When diluting the standard or samples, <u>change tips</u> between each standard or sample dilution. When loading reagents in the assay microplate, <u>change tips</u> between each standard or sample addition and after each set of reagents.
- When reagents are added to the microplate, make sure the liquids are at the bottom of the well.
- Small volumes may be prone to evaporation. It is recommended to cover microplates with TopSeal-A Adhesive Sealing Films to reduce evaporation during incubation. Microplates can be read with the TopSeal-A Film.
- The AlphaLISA signal is detected with an EnVision Multilabel Reader equipped with the Alpha option using the AlphaScreen standard settings (e.g. Total Measurement Time: 550 ms, Laser 680 nm Excitation Time: 180 ms, Mirror: D640as, Emission Filter: M570w, Center Wavelength 570 nm, Bandwidth 100 nm, Transmittance 75%).
- AlphaLISA signal will vary with temperature and incubation time. For consistent results, identical incubation times and temperature should be used for each plate.
- The standard curves shown in this technical data sheet are provided for information only. A standard curve must be generated for each experiment.

Assay Procedure

IMPORTANT: PLEASE READ THE RECOMMENDATIONS BELOW BEFORE USE

- The protocol described below is an example for generating one standard curve in a 50 µL final assay volume (48 wells, triplicate determinations). The protocols also include testing samples in 452 wells. If a different amount of samples are tested, the volumes of all reagents have to be adjusted accordingly, as shown in the table below. These calculations do not include excess reagent to account for losses during transfer of solutions or dead volumes.
- The standard dilution protocol is provided for information only. As needed, the number of replicates or the range of concentrations covered can be modified.
- Use of four background points in triplicate (12 wells) is recommended when LDL/LLOQ is calculated. One background point in triplicate (3 wells) can be used when LDL/LLOQ is not calculated.



		Volume				
Format	# of data points	Final	Sample	AlphaLISA beads / Biotin Antibody MIX	SA- Donor beads	Plate recommendation
AL532HV	100	100 μL	10 μL	40 μL	50 μL	White OptiPlate-96 (cat # 6005290) White ½ AreaPlate-96 (cat # 6005560)
	250	100 μL	10 μL	40 μL	50 µL	White OptiPlate-96 (cat # 6005290) White ½ AreaPlate-96 (cat # 6005560)
A1 5200	500	50 μL	5 µL	20 μL	25 μL	White ½ AreaPlate-96 (cat # 6005560) White OptiPlate-384 (cat # 6007290) Light gray AlphaPlate™-384 (cat # 6005350)
AL532C	1 250	20 μL	2 µL	8 µL	10 μL	Light gray AlphaPlate-384 (cat # 6005350) ProxiPlate™-384 Plus (cat # 6008280) White OptiPlate-384 (cat # 6007290)
	2 500	10 μL	1 μL	4 µL	5 μL	Light gray AlphaPlate-1536 (cat # 6004350)
	5 000	50 μL	5 µL	20 μL	25 μL	White ½ AreaPlate-96 (cat # 6005560) White OptiPlate-384 (cat # 6007290) Light gray AlphaPlate-384 (cat # 6005350)
AL532F	12 500	20 μL	2 µL	8 µL	10 μL	Light gray AlphaPlate-384 (cat # 6005350) ProxiPlate-384 Plus (cat # 6008280) White OptiPlate-384 (cat # 6007290)
	25 000	10 μL	1 µL	4 μL	5 μL	Light gray AlphaPlate-1536 (cat # 6004350)



The 3-step high sensitivity protocol described below is for 500 assay points including one standard curve (48 wells) and samples (452 wells).

- 1) Preparation of 1X AlphaLISA Bovine Immunoassay Buffer:
 - Add 10 mL of 10X AlphaLISA Bovine Immunoassay Buffer to 90 mL H₂O.
- 2) Preparation of blgG2 analyte standard dilutions:
 - Bovine IgG2 analyte is provided at 3 μg in lyophilized form. Reconstitute with 100 μL MiliQ H₂O to create a 30 μg/mL solution. The first point of the curve is 3 μg/mL so a 10 fold dilution is required. Prepare standard dilutions as follows (change tip between each standard dilution):

Tube	Vol. of	Vol. of diluent (μL) *	[Bovine IgG2] in standard curve		
	Bovine IgG2 (μL)	unuent (µL)	(g/mL in 5 µL)	(pg/mL in 5 µL)	
Α	10 μL of provided IgG2	90	3.00E-06	3000000	
В	60 μL of tube A	120	1.00E-06	1000000	
С	60 μL of tube B	140	3.00E-07	300000	
D	60 μL of tube C	120	1.00E-07	100000	
Е	60 μL of tube D	140	3.00E-08	30000	
F	60 μL of tube E	120	1.00E-08	10000	
G	60 μL of tube F	140	3.00E-09	3000	
Н	60 μL of tube G	120	1.00E-09	1000	
I	60 μL of tube H	140	3.00E-10	300	
J	60 μL of tube I	120	1.00E-10	100	
K	60 μL of tube J	140	3.00E-11	30	
L	60 μL of tube K	120	1.00E-11	10	
M ** (background)	0	100	0	0	
N ** (background)	0	100	0	0	
O ** (background)	0	100	0	0	
P ** (background)	0	100	0	0	

- * Dilute standards in diluent (e.g. 1X AlphaLISA Bovine Immunoassay Buffer).
 - At low concentrations of analyte, a significant amount of analyte can bind to the vial. Therefore, load the analyte standard dilutions in the assay microplate within 60 minutes of preparation.
- ** Four background points in triplicate (12 wells) are used when LDL is calculated. If LDL does not need to be calculated, one background point in triplicate can be used (3 wells).
- 3) Preparation of 5X AlphaLISA Anti-bovine IgG2 Acceptor beads (50 μg /mL) Add 60 μL of 5 mg/mL AlphaLISA Anti- bovine IgG2 acceptor beads to 5970 μL of AlphaLISA Immunoassay Buffer. Prepare just before use.
- 4) Preparation of 5X biotinylated anti- bovine IgG2 antibody (10 nM): Add 120 μL of 500nM biotinylated anti- bovine IgG2 antibody to 5940 μL of 1X AlphaLISA Immunoassay Buffer. Prepare just before use.
- 5) <u>Preparation of 2X Streptavidin (SA) Donor beads</u> (80 μg/mL): Keep the beads under subdued laboratory lighting. **Prepare just before use.**
 - Add 200 μ L of 5 mg/mL SA-Donor beads to 12 300 μ L of 1X AlphaLISA Bovine Immunoassay Buffer. **Prepare just before use.**



6) In a 96- or 384-well microplate:

Add 5 µL of each IgG2 analyte standard dilution or 5 µL of sample

Add 10 µL of a 5X AlphaLISA Anti-bovine IgG2 Acceptor beads (10 µg/mL final)

Incubate 30 minutes at 23 °C

Add 10 µL of a 5X Biotinylated Anti-bovine IgG2 Antibody (2 nM final)

Incubate 60 minutes at 23°C

Add 25 µL of 2X SA-Donor beads (40 µg/mL final)

Incubate 30 minutes at 23°C in the dark

Read using EnVision-Alpha Reader

Data Analysis

Calculate the average count value for the background wells.



- Generate a standard curve by plotting the AlphaLISA counts versus the concentration of analyte. A log scale can be used for either or both axes. No additional data transformation is required.
- Analyze data according to a nonlinear regression using the 4-parameter logistic equation (sigmoidal dose-response curve with variable slope) and a 1/Y² data weighting (the values at maximal concentrations of analyte after the hook point should be removed for correct analysis).
- The LDL is calculated by interpolating the average background counts (12 wells without analyte) + 3 x standard deviation value (average background counts + (3xSD)) on the standard curve.
- The LLOQ as measured here is calculated by interpolating the average background counts (12 wells without analyte) + 10 x standard deviation value (average background counts + (10xSD)) on the standard curve. Alternatively, the true LLOQ can be determined by spiking known concentrations of analyte in the matrix and measuring the percent recovery, and then determining the minimal amount of spiked analyte that can be quantified within a given limit (usually +/- 20% or 30% of the real concentration).
- Read from the standard curve the concentration of analyte contained in the samples.
- If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

Assay Performance Characteristics

AlphaLISA assay performance described below was determined using the Quick Protocol.

Assay Sensitivity and Precision:

The LDL and LLOQ were calculated as described above. The values correspond to the lowest concentration of analyte that can be detected in a volume of 5 µL using the recommended assay conditions. LDL, LLOQ, EC50, maximum, and minimum counts in table below were summarized from 30 independent assays.

Parameters	Average	Minimum	Maximum
LDL ng/mL	2.06	0.62	4.66
LLOQ ng/mL	5.67	1.91	10.91
EC ₅₀ ng/mL	179	82	299
Maximum Counts	169132	90691	420068
Minimum Counts	3600	1723	6455
Intra-Asaay CV (%)	4.7	1.4	10.5
Inter Assay CV (%)	6.4	NA	NA

* Note that LDL/ LLOQ can be decreased (i.e. sensitivity increased) by increasing the volume of analyte in the assay (e.g. use 10 μ L of analyte in a final assay volume of 50 μ L).

Specificity

Cross-reactivity of the AlphaLISA bovine IgG2 Kit was tested using the following proteins at 0.1 µg/mL in AlphaLISA Bovine Immunoassay Buffer. Reactivity to bovine IgG2 is 100%.

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Proteins	Cross Reactivity (%)
Bovine lgG1	0.00
Bovine lgA	0.43
Bovine IgM	2.25
Human lgG	0.00
Horse IgG	0.00
Swine lgG	0.00
Goat lgG	0.00

Troubleshooting Guide

You will find detailed recommendations for common situations you might encounter with your AlphaLISA Assay kit at:

http://www.perkinelmer.com/in/resources/technicalresources/applicationsupportknowledgebase/alphalisa-alphascreen-no-washassays/alpha_troubleshoot.xhtml

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