

LANCE *Ultra* SET7/9 Histone H3-Lysine N-methyltransferase Assay

U-TRF #35

LANCE® *Ultra*

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This LANCE® *Ultra* immunodetection assay measures the mono-methylation of a biotinylated Histone H3 (1-21) peptide at lysine 4.

Europium-anti-methyl-Histone H3 Lysine 4 (H3K4me1-2) Antibody

- TRF0402-D: 10 µg, 1,562 assay points*
- TRF0402-M: 100 µg, 15,625 assay points*

*40 fmol/assay point

Peptidic Substrate Sequence:

ARTKQTARKSTGGKAPRKQLA-GG-K(BIOTIN)-NH₂

LANCE *Ultra* Assays

LANCE *Ultra* time-resolved fluorescence resonance energy transfer (TR-FRET) assays use a proprietary europium chelate donor dye, W1024 (Eu), together with *ULight*™, a small molecular weight acceptor dye with a red-shifted fluorescent emission.

In this technical note, we present the optimization of an epigenetic enzymatic assay using a biotinylated histone H3-derived peptide as substrate. The modified peptide is captured by the Eu-labeled antibody (Eu-Ab) and *ULight*-Streptavidin (SA) which bring the Eu donor and *ULight* acceptor dye molecules into close proximity. Upon irradiation at 320 or 340 nm, the energy from the Eu donor is transferred to the *ULight* acceptor dye which, in turn, generates light at 665 nm. The intensity of the light emission is proportional to the level of biotinylated substrate modification.

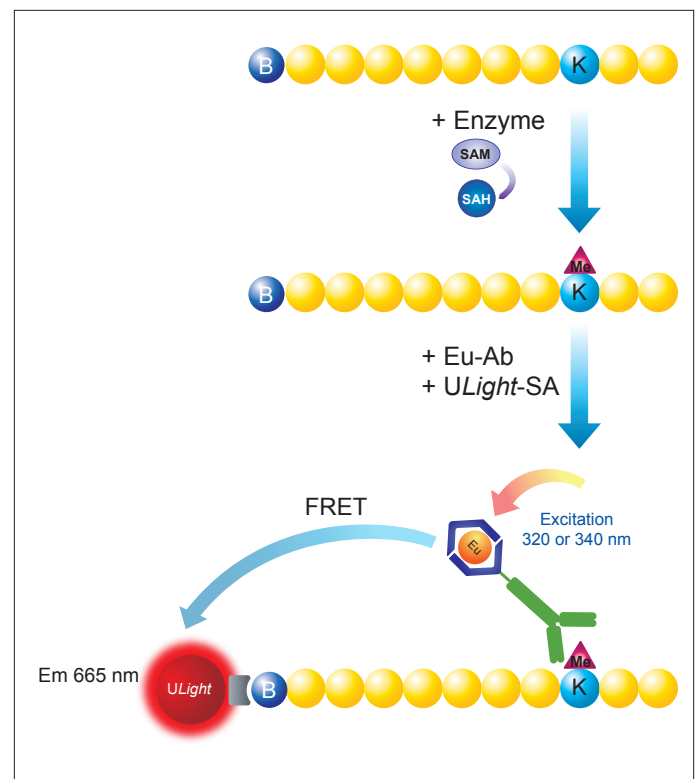


Figure 1. Schematic representation of the LANCE *Ultra* detection of a modified histone peptide.

Development of a SET7/9 Histone H3-Lysine N-methyltransferase Assay

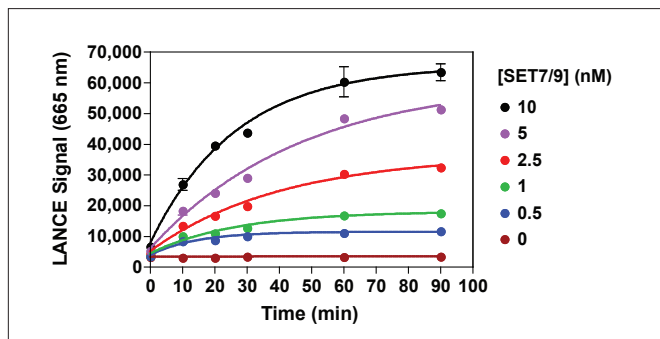
Reagents needed for the assay:

Europium-anti-methyl-Histone H3 Lysine 4 (H3K4me1-2)	PerkinElmer # TRF0402
LANCE <i>Ultra ULight</i> -Streptavidin	PerkinElmer # TRF0102
Histone H3 (1-21) peptide, biotinylated	AnaSpec # 61702
LANCE Detection Buffer, 10X	PerkinElmer # CR97-100
SET7/9 (human), recombinant	Enzo # ALX-201-178-C100
White opaque OptiPlate™-384	PerkinElmer # 6007299
TopSeal™-A films	PerkinElmer # 6005185
S-(5'-Adenosyl)-L-methionine chloride (SAM)	Sigma # A7007
Sinefungin	Sigma # S8559
S-(5'-Adenosyl)-L-homocysteine (SAH)	Sigma # A9384

SAM is prepared at 30 mM in 5 mM H₂SO₄/10% ethanol (v/v) in H₂O, aliquoted and stored at -80 °C.

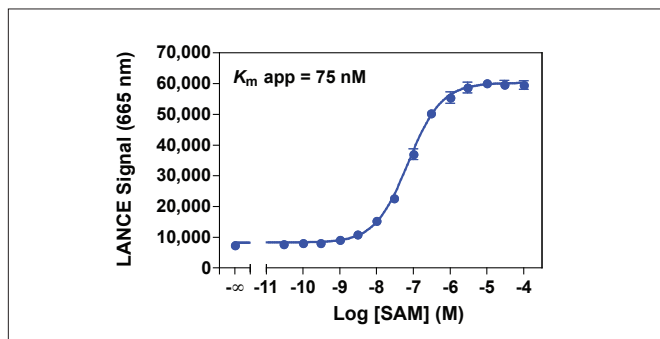
Assay Buffer: 50 mM Tris-HCl, pH 8.8, 5 mM MgCl₂, 1 mM DTT, 0.01% Tween-20

Experiment 1: Enzyme Titration and Time-Course



Enzymatic progress curves were performed by incubating SET7/9 at concentrations ranging from 0.5 to 10 nM with 200 nM biotinylated H3 (1-21) peptide substrate and 300 μM SAM. Detection Mix was added to stop the reactions at the indicated times and signal was read after 60 min. A 60 min reaction time using 5 nM enzyme was selected for all subsequent experiments.

Experiment 2: SAM Titration



Serial dilutions of SAM ranging from 30 pM to 100 μM were added to 5 nM SET7/9 and 200 nM biotinylated H3 (1-21) peptide substrate. A 300 nM SAM concentration was selected for subsequent experiments.

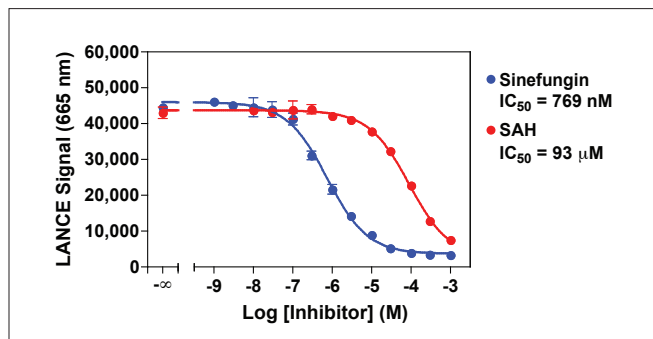
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Standard Protocol

- Dilute SET7/9 enzyme, SAM, inhibitors and biotinylated peptide substrate in Assay Buffer just before use.
- Add to the wells of a white Optiplate-384:
 - 5 μL of inhibitor (2X) or Assay Buffer
 - 2.5 μL of enzyme (4X)
 - 2.5 μL of biotinylated Histone H3 (1-21) peptide/SAM mix (4X). For SAM titration, add SAM dilutions independently of substrate.
- Cover the plate with TopSeal-A film and incubate at room temperature (RT).
- Prepare Detection Mix* by diluting the Eu-Ab to 4 nM and *ULight*-Streptavidin to 100 nM in 1X LANCE Detection Buffer (final concentrations of 2 nM and 50 nM respectively, in 20 μL total assay volume).
- Add 10 μL of Detection Mix.
- Cover with TopSeal-A film and incubate for 60 min at RT.
- Remove the TopSeal-A film and read signal with EnVision® Multilabel Reader in TR-FRET mode (excitation at 320 or 340 nm & emission at 665 nm).

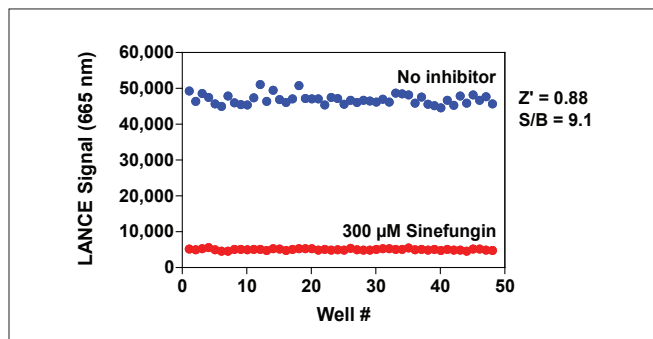
* Under these experimental conditions, addition of the Detection Mix stops SET7/9 enzymatic reaction.

Experiment 3: Enzyme Inhibition



Serial dilutions of sinefungin ranging from 1 nM to 1 mM and SAH ranging from 10 nM to 1 mM were pre-incubated for 10 min with 5 nM SET7/9. Enzymatic reactions were initiated by the addition of 200 nM biotinylated H3 (1-21) peptide substrate plus 300 nM SAM. Enzymatic reactions contain 1% DMSO.

Experiment 4: Z'-factor Determination



SET7/9 (5 nM) was pre-incubated with or without 300 μM sinefungin for 10 min. Enzymatic reactions were initiated by the addition of 200 nM biotinylated H3 (1-21) peptide substrate plus 300 nM SAM. Enzymatic reactions contain 1% DMSO.