

c-TAK1 Kinase Assay

ULight-Cdc25C (Ser216) Peptide and Europium-anti-phospho-(Ser) 14-3-3 binding motif 4E2 Antibody

Two LANCE Ultra companion products!

ULight™-Cdc25C (Ser216) Peptide:

- TRF0123-D: 1 nmole, 1,000* assay points
- TRF0123-M: 10 nmoles, 10,000* assay points

*1 pmol/assay point

PEPTIDE SEQUENCE:

CSRSGLYRSPSMPENLNRPRL

Synthetic peptide derived from residues 207-226 of human M-phase inducer phosphatase 3 (Cdc25C); phosphorylation site: Ser216.

VALIDATED FOR KINASE: c-TAK1

POTENTIAL SUBSTRATE FOR KINASES: CHK1, CHK2

Europium-anti-phospho-(Ser) 14-3-3 binding motif 4E2 Antibody:

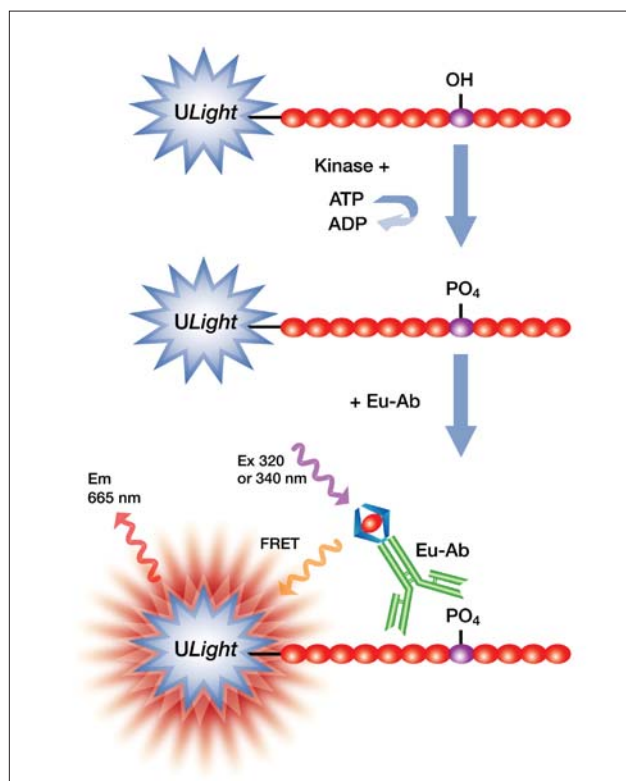
- AD0192: 10 µg, 1,562* assay points

*40 fmol/assay point

RECOGNIZED MOTIF:

(R/K)XXpSXP

Europium-labeled mouse monoclonal antibody.



LANCE Ultra Kinase Assays

LANCE® Ultra time-resolved fluorescence resonance energy transfer (TR-FRET) assays use a proprietary europium chelate donor dye, W-1024 (Eu), with ULight, an innovative small molecular weight acceptor dye with a red-shifted fluorescent emission. In kinase assays, the binding of a Eu-labeled anti-phospho-substrate antibody to the phosphorylated ULight-labeled substrate brings donor and acceptor molecules into close proximity.

After irradiation of the kinase reaction at 320 or 340 nm, the energy from the Eu donor is transferred to the ULight acceptor dye which, in turn, generates light at 665 nm. The intensity of the light emission is proportional to the level of ULight-substrate phosphorylation.

Development of a c-TAK1 Kinase Assay

Additional reagents

C-TAK1, active	Upstate #14-375
LANCE Detection Buffer, 10X	PerkinElmer # CR97-100
OptiPlate™-384, white	PerkinElmer # 6007299
TopSeal™-A	PerkinElmer # 6005185
Kinase Buffer: 50 mM HEPES pH 7.5, 1 mM EGTA, 10 mM MgCl ₂ , 2 mM DTT and 0.01% Tween-20.	

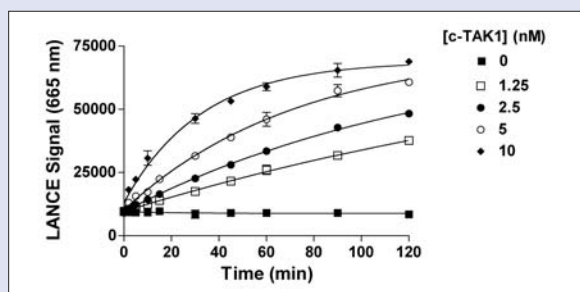
Suggested procedure

- Dilute the c-TAK1 kinase, ATP, inhibitors and *ULight*-Cdc25C (Ser216) peptide in Kinase Buffer.
- Prepare a 4X Detection Mix by diluting the Eu-anti-phospho (Ser) 14-3-3 binding motif 4E2 antibody to 8 nM in 1X LANCE Detection Buffer.
- Add to the wells of a white Optiplate-384:
 - 5 μ L of c-TAK1 enzyme
 - 2.5 μ L of inhibitor or Kinase Buffer
 - 2.5 μ L of *ULight*-Cdc25C (Ser216) peptide/ ATP mix (for ATP titration, ATP dilutions are added separately in Kinase Buffer).

- Cover the plate with TopSeal-A and incubate at room temperature (RT).
- Stop kinase reactions by adding 5 μ L of 40 mM EDTA prepared in 1X Detection Buffer (Stop Solution). Leave for 5 min at RT.
- Add 5 μ L of 4X Detection Mix (Eu-anti-phospho-(Ser) 14-3-3 binding motif 4E2 Antibody at a final concentration of 2 nM).
- Cover with TopSeal-A and incubate for 1 h at RT.
- Remove TopSeal-A and read signal with the EnVision[®] Multilabel Reader in TR-FRET mode (excitation at 320 nm and emission at 665 nm).

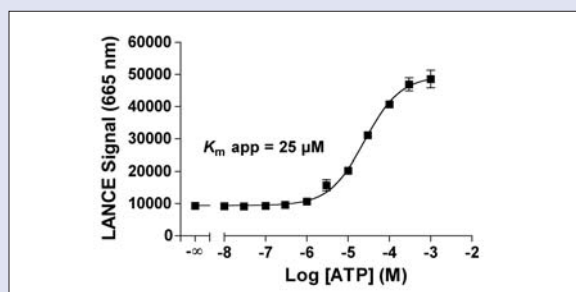
NOTE: Eu-labeled antibodies and EDTA can be premixed before use as a 2X concentrated Stop Solution/Detection Mix to minimize the number of liquid handling steps.

Experiment 1: Enzymatic Time Course



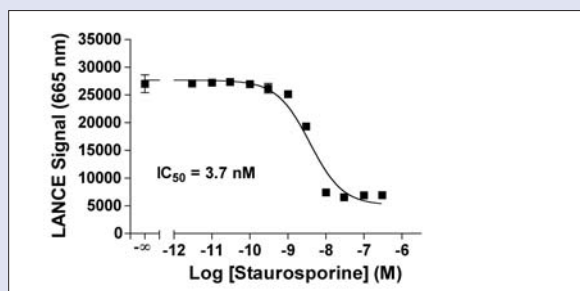
c-TAK1 enzyme was incubated at concentrations ranging from 1.25 to 10 nM with 100 nM *ULight*-Cdc25C (Ser216) peptide and 50 μ M ATP. Kinase reactions were terminated after 0 to 120 min by the addition of EDTA.

Experiment 2: ATP Titration



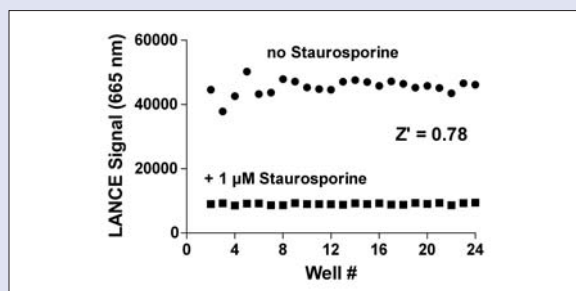
Serial dilutions of ATP ranging from 10 nM to 1 mM were added to 5 nM c-TAK1 and 100 nM *ULight*-Cdc25C (Ser216) peptide. Kinase reactions were terminated after 90 min by the addition of EDTA.

Experiment 3: Enzyme Inhibition Curve



Serial dilutions of staurosporine ranging from 3 pM to 300 nM (final concentrations in 2% DMSO) were incubated with 5 nM c-TAK1, 100 nM *ULight*-Cdc25C (Ser216) peptide and 100 μ M ATP. Kinase reactions were terminated after 45 min by the addition of EDTA.

Experiment 4: Z'-factor Determination



c-TAK1 enzyme at 5 nM was incubated with 100 nM *ULight*-Cdc25C (Ser216) peptide and 100 μ M ATP with or without 1 μ M staurosporine (final concentrations in 2% DMSO). Kinase reactions were terminated after 45 min by the addition of EDTA.