

U-TRF #27

# LANCE *Ultra* EphA4 Kinase Assay

LANCE® *Ultra* TR-FRET Technology

PerkinElmer, Inc.  
Montreal, QC  
Canada H3J 1R4

This LANCE *Ultra* kinase assay measures the phosphorylation of the *ULight*<sup>™</sup>-TK peptide substrate.

- TRF0127-D: 0.5 nmole, 1,000 assay points\*
- TRF0127-M: 5 moles, 10,000 assay points\*

\*0.5 pmol/assay point

### Europium-anti-phosphotyrosine (PT66):

- AD0068: 50 µg, 7,800 assay points\*
- AD0069: 1 mg, 156,000 assay points\*

\*40 fmol/assay point

### Recognized Motif:

Phosphorylated tyrosine residues

### LANCE *Ultra* Assays

LANCE *Ultra* time-resolved fluorescence resonance energy transfer (TR-FRET) assays use a proprietary europium chelate donor dye, W1024 (Eu), together with *ULight*<sup>™</sup>, a small molecular weight acceptor dye with a red-shifted fluorescent emission.

In this technical note, we present the optimization of a EphA4 kinase assay using a *ULight*-labeled peptide substrate. The binding of the Eu-labeled PT66 antibody to the phosphorylated *ULight*-TK peptide substrate brings the Eu donor and *ULight* acceptor dye molecules into close proximity. Upon irradiation at 320 or 340 nm, the energy from the Eu donor is transferred to the *ULight* acceptor dye which, in turn, generates light at 665 nm. The intensity of the light emission is proportional to the level of *ULight* substrate phosphorylation.

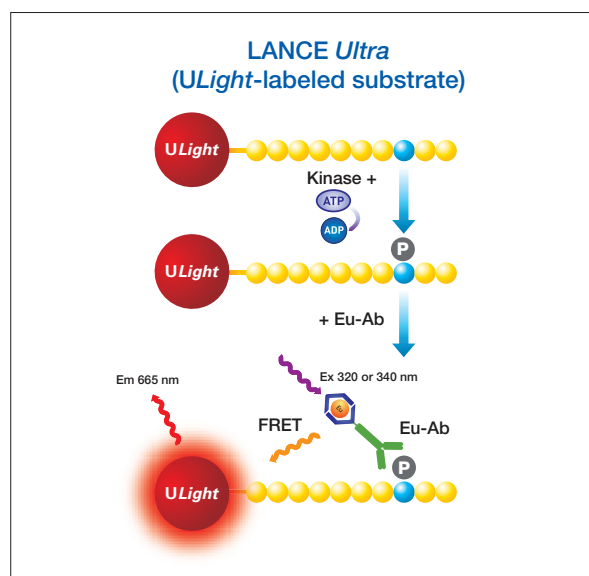


Figure 1. Schematic representation of the LANCE *Ultra* detection of a phosphorylated peptide substrate.

## Development of an EphA4 Kinase Assay

### Reagents needed for this assay:

Europium-labeled anti-phosphotyrosine Antibody	PerkinElmer # AD0068/AD0069
ULight-TK Peptide	PerkinElmer # TRF0127
EphA4	Carna # 08-123
LANCE Detection Buffer, 10X	PerkinElmer # CR97-100
White opaque OptiPlate™-384	PerkinElmer # 6007290
TopSeal™-A Film	PerkinElmer # 6050195

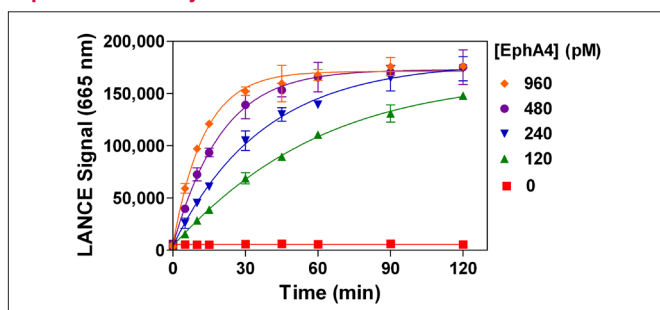
Kinase Buffer: 50 mM HEPES pH 7.5, 1 mM EGTA, 10 mM MgCl<sub>2</sub>, 2 mM DTT and 0.01% Tween-20.

### Standard Protocol

- Dilute the EphA4 enzyme, ATP, inhibitors and ULight-TK Peptide in Kinase Buffer.
- Prepare a 4X Detection Mix by diluting the Eu-anti-phosphotyrosine Antibody to 8 nM in 1X LANCE Detection Buffer.

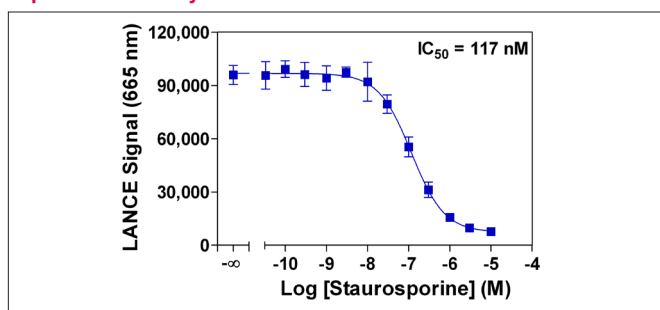
## Results

### Experiment 1: Enzyme Titration and Time-Course



Enzymatic progress curves were produced by incubating the EphA4 enzyme at concentrations ranging from 120 to 960 pM with 50 nM ULight-TK Peptide and 200 μM ATP. Kinase reactions were terminated at the indicated times by the addition of EDTA. Detection mix was added and signal read after 60 minutes.

### Experiment 3: Enzyme Inhibition Curve



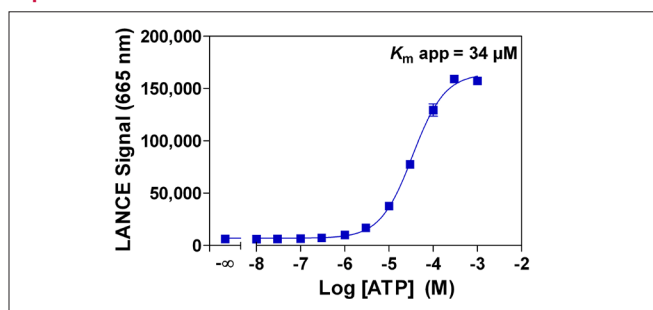
Increasing concentrations of Staurosporine ranging from 30 pM to 10 μM (final concentrations in 2% DMSO) were incubated with 120 pM EphA4 enzyme, 50 nM ULight-TK Peptide and 35 μM ATP. Kinase reactions were terminated after 60 min by the addition of EDTA.

PerkinElmer, Inc.  
940 Winter Street  
Waltham, MA 02451 USA  
P: (800) 762-4000 or  
(+1) 203-925-4602  
[www.perkinelmer.com](http://www.perkinelmer.com)

- Add to the wells of a white OptiPlate-384:
  - 5 μL of EphA4 enzyme
  - 2.5 μL of inhibitor or Kinase Buffer
  - 2.5 μL of ULight-TK Peptide/ATP mix (for ATP titration, ATP dilutions are added separately in Kinase Buffer).
- Cover the plate with TopSeal-A film and incubate at RT.
- Stop kinase reactions by adding 5 μL of 40 mM EDTA prepared in 1X LANCE Detection Buffer (Stop Solution). Leave for 5 min at RT.
- Add 5 μL of Detection Mix (Eu-anti-phosphotyrosine Antibody at a final concentration of 2 nM).
- Cover with TopSeal-A film and incubate 1 h at RT.
- Remove the TopSeal-A film and read signal with the EnVision® Multilabel Plate Reader in TR-FRET mode (excitation at 320 or 340 nm and emission at 665 nm).

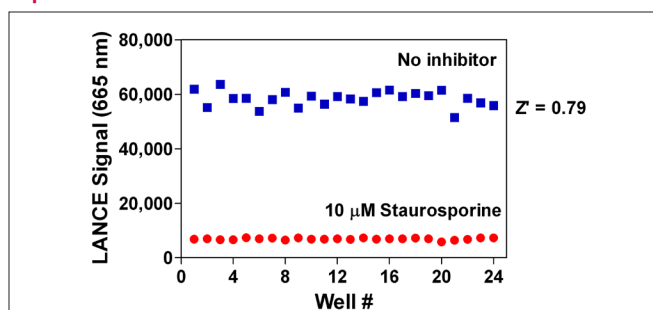
**NOTE:** Eu-labeled antibodies and EDTA can be premixed just before use as a 2X concentrated Stop Solution/Detection mix to minimize the number of liquid handling steps.

### Experiment 2: ATP Titration



Increasing concentrations of ATP ranging from 10 nM to 1 mM were incubated with 120 pM EphA4 enzyme and 50 nM of ULight-TK Peptide. Kinase reactions were terminated after 60 min by the addition of EDTA.

### Experiment 4: Z'-factor Determination



EphA4 enzyme at 120 pM was incubated with 50 nM ULight-TK Peptide in kinase assay buffer with 35 μM ATP, and with or without 10 μM staurosporine (final concentrations in 2% DMSO). Kinase reactions were terminated after 60 min by the addition of EDTA. A Z' value of 0.79 indicates that the assay is very robust and can be used for HTS.



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