

LANCE *Ultra* ERK1 Assay

Using *ULight*-Myelin Basic Protein (MBP) Peptide & Europium-Anti-Phospho-MBP Peptide Antibody

Two LANCE® *Ultra* companion products—two convenient sizes!

ULight™-MBP Peptide:

- **TRF0109-D: 0.5 nmole, 1,000 assay points***
- **TRF0109-M: 5 nmoles, 10,000 assay points***
*0.5 pmol/assay point
- **PEPTIDE SEQUENCE:**
CFFKNIVTPRTPPPSQGK-amide
 - Synthetic peptide derived from human myelin basic protein (MBP)
 - Phosphorylation site: THR232*
- *corresponds to Thr98 in other MBP isoforms or species
- **VALIDATED FOR KINASES:** ERK1, ERK2, CDK3/CycE
- **POTENTIAL SUBSTRATE FOR KINASES:** s6K, p43, p38

Europium-anti-phospho-Myelin Basic Protein (Thr232) antibody:

- **TRF0201-D: 10 µg, 1,562 assay points***
- **TRF0201-M: 100 µg, 15,625 assay points***
*40 fmol/assay point
- **RECOGNIZED MOTIF:** FFKNIVTPR**p**TPPPSQGK
- Europium-labeled mouse monoclonal antibody recognizing phospho-Thr232 in human Myelin Basic Protein (Swiss-Prot: P02686)

LANCE *Ultra* Kinase Assays

LANCE® *Ultra* time-resolved fluorescence resonance energy transfer (TR-FRET) assays use a proprietary europium chelate donor dye, W-1024 (Eu), with *ULight*, a new innovative small molecular weight acceptor dye with a red-shifted fluorescent emission. In kinase assays, the binding of an Eu-labeled anti-phospho-substrate antibody to phosphorylated *ULight*-labeled substrates brings donor and acceptor molecules into close proximity.

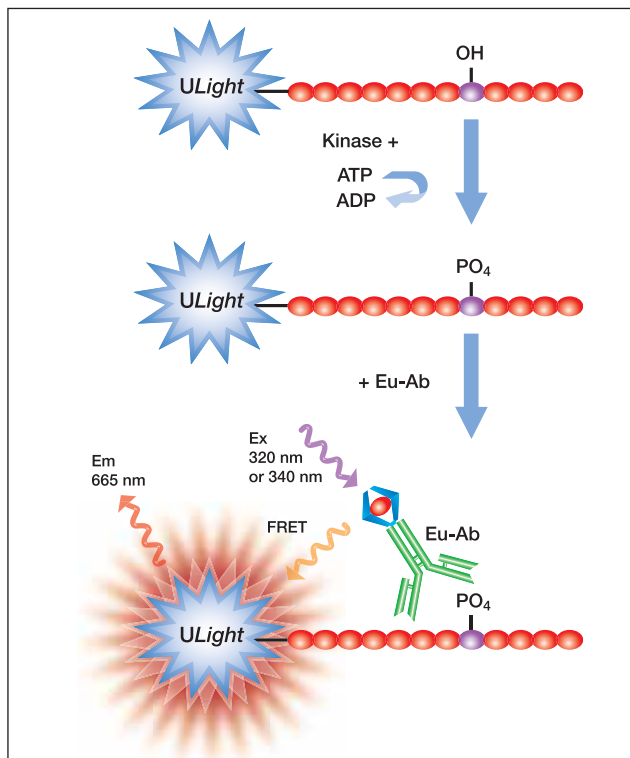
After irradiation of the kinase reaction at 320 nm, the energy from the Eu donor is transferred to the *ULight* acceptor which, in turn, generates light at 665 nm. The intensity of the light emission is proportional to the level of *ULight*-substrate phosphorylation.

Development of a ERK1 Kinase Assay

Additional Reagents

ERK1, active	Upstate # 14-439
LANCE Detection Buffer, 10X	PerkinElmer # CR97-100
OptiPlate™-384, white	PerkinElmer # 6007299
TopSeal-A™	PerkinElmer # 6005185

Kinase Buffer: 50 mM Tris-HCl, pH 7.5, 1 mM EGTA, 10 mM MgCl₂, 2 mM DTT and 0.01% Tween-20



Suggested Procedure

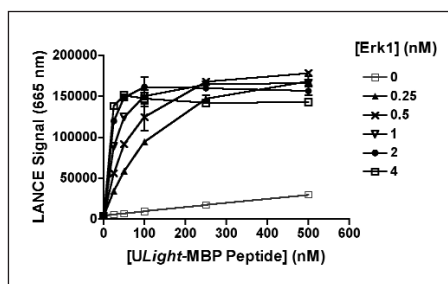
- Dilute kinase, ATP, inhibitors and *ULight*-MBP in Kinase Buffer.
- Dilute antibody (Ab) in LANCE Detection Buffer to 8 nM.
- Add to the wells of a white OptiPlate-384:
 - 5 μ L of ERK1 enzyme,
 - 2.5 μ L of inhibitor or Kinase Buffer,
 - 2.5 μ L of *ULight*-MBP/ATP mix (for ATP titration, ATP dilutions are added separately in Kinase Buffer).
- Incubate enzymatic reactions at room temperature (RT).
- Stop the reaction by adding 5 μ L of 40 mM EDTA in Detection Buffer. Leave 5 min at RT.
- Add 5 μ L of the antibody dilution (2 nM final concentration).
- Incubate for 1 h at RT.
- Remove TopSeal-A and read signal with the EnVision™ Multilabel Reader in TR-FRET mode (excitation at 320 nm and emission at 665 nm).

Better ERK1 Kinase Assays with a Better Technology—LANCE Ultra

For more information about LANCE Ultra, please visit www.perkinelmer.com/lanceultra or contact your local PerkinElmer Sales Representative. Learn more about our comprehensive range of reagents and consumables for drug discovery by visiting www.perkinelmer.com/drugdiscovery.

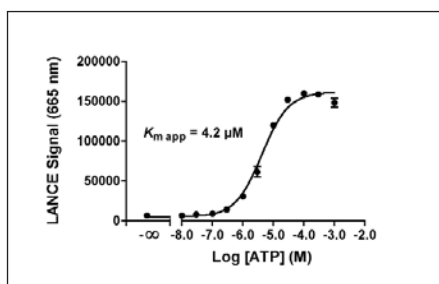
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Experiment 1: Enzymatic Time Course



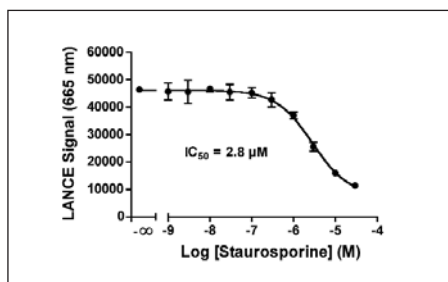
ERK1 enzyme ranging from 0.25 to 4 nM was incubated with the *ULight*-MBP substrate ranging from 25 to 500 nM in kinase assay buffer supplemented with 20 μ M ATP. Reactions were terminated after 90 min by the addition of EDTA.

Experiment 2: ATP Titration



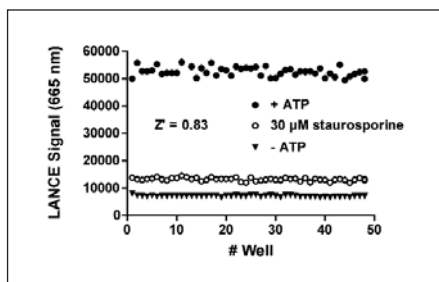
Serial dilutions of ATP ranging from 10 nM to 1 μ M were added to 1 nM of ERK1 kinase and 50 nM of *ULight*-MBP substrate. Kinase reactions were terminated after 90 min by the addition of EDTA.

Experiment 3: Enzyme Inhibition Curve



Serial dilutions of staurosporine ranging from 1 nM to 30 μ M (final concentrations in 2% DMSO) were pre-incubated for 5 min with the ERK1 enzyme (1 nM final concentration). Then 50 nM *ULight*-MBP and 4 μ M ATP were added. Kinase reactions were terminated after 60 min by the addition of EDTA.

Experiment 4: Z'-factor Determination



The ERK1 enzyme at 1 nM was incubated with 50 nM *ULight*-MBP substrate in kinase assay buffer with 4 μ M ATP, 30 μ M staurosporine and ATP, or without ATP. 2% DMSO was included. Reactions were terminated after 60 min by the addition of EDTA.

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