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Human Fibronectin (hFN) AlphaLISA Detection Kit

Product No.: AL3056C/F

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Product Information

Application:	This kit is designed for the quantitative determination of fibronectin (FN) in human serum and cell culture media (DMEM and RPMI) using a homogeneous AlphaLISA assay (no wash steps). The assay shows negligible cross-reactivity (<1%) with human collagen Type I, collagen Type III, vitronectin, and hFN fragment III.1, and with mouse, rat, and bovine fibronectin.
Sensitivity:	Lower Detection Limit (LDL): 242 pg/mL Lower Limit of Quantification (LLOQ): 785 pg/mL EC ₅₀ : 308 ng/mL
Dynamic range:	242 – 3 000 000 pg/mL

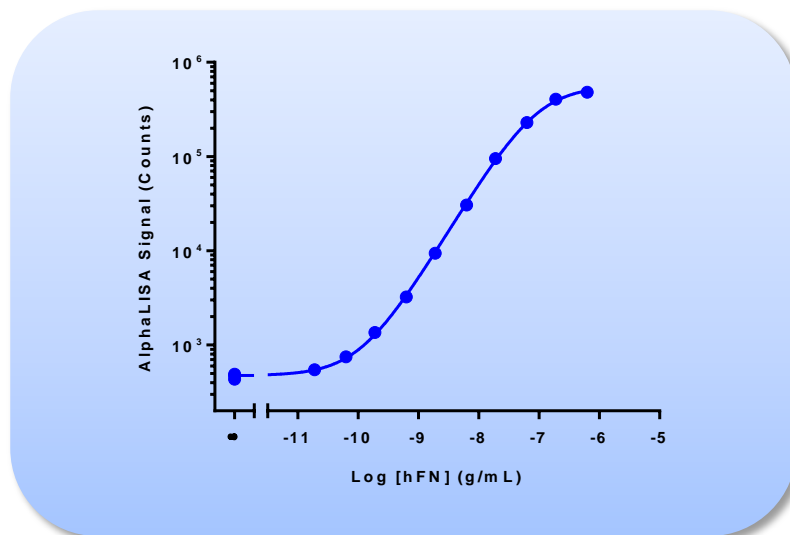


Figure 1. Typical sensitivity curve in AlphaLISA Immunoassay Buffer. The data was generated using a white Optiplate™-384 microplate and the EnVision® Multilabel Plate Reader 2103 with Alpha option.

Storage:	Store kit in the dark at +4°C. For reconstituted analyte aliquot and store at -20 °C. Avoid freeze-thaw cycles.
Stability:	This kit is stable for at least 6 months from the manufacturing date when stored in its original packaging and the recommended storage conditions.

Quality Control

Lot to lot consistency is confirmed in an AlphaLISA assay. Maximum and minimum signals, EC₅₀ and LDL were measured on the EnVision Multilabel Plate Reader with Alpha option using the protocol described in this technical data sheet. We certify that these results meet our quality release criteria. Maximum counts may vary between bead lots and the instrument used, with no impact on LDL measurement.

Analyte of Interest

Fibronectin is a large glycoprotein of the extracellular matrix that exists in two forms: insoluble (cellular) and as a soluble plasma protein. The primary functions of fibronectin are its involvement in cell adhesion, growth, migration, and differentiation. Fibronectin is known to contribute to the wound healing process where it will accumulate at the site of injury to help form blood clots and aid in the development of new tissues. Fibronectin levels in serum and plasma have been identified to increase significantly after major traumas. Further, Fibronectin levels have been observed to be significantly altered in disease states of certain types of cancer and also diabetes mellitus. This AlphaLISA kit has been designed to detect and quantify Human Fibronectin in human serum and cell culture supernatants.

Description of the AlphaLISA Assay

AlphaLISA technology allows the detection of molecules of interest in buffer, cell culture media, serum and plasma in a highly sensitive, quantitative, reproducible and user-friendly mode. In an AlphaLISA assay, a Biotinylated Anti-hFN Antibody binds to the Streptavidin-coated Alpha Donor beads, while another Anti-hFN antibody is conjugated to AlphaLISA Acceptor beads. In the presence of the human fibronectin, the beads come into close proximity. The excitation of the Donor beads provokes the release of singlet oxygen molecules that triggers a cascade of energy transfer in the Acceptor beads, resulting in a sharp peak of light emission at 615 nm (Figure 2).

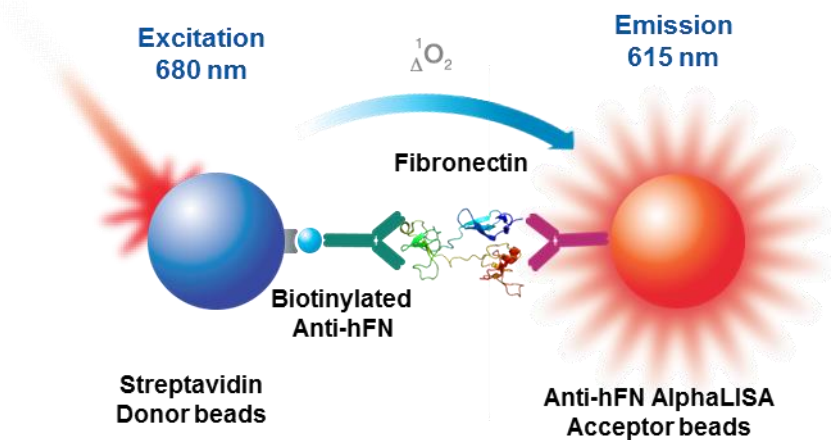


Figure 2. AlphaLISA Assay Principle.

Precautions

- The Alpha Donor beads are light-sensitive. All the other assay reagents can be used under normal light conditions. All Alpha assays using the Donor beads should be performed under subdued laboratory lighting (< 100 lux). Green filters (LEE 090 filters (preferred) or Roscolux filters #389 from Rosco) can be applied to light fixtures.
- Take precautionary measures to avoid contamination of the reagent solutions.
- The Biotinylated Anti-hFN Antibody contains sodium azide. Contact with skin or inhalation should be avoided.

Kit Content: Reagents and Materials

Kit components	AL3056HV (100 assay points***)	AL3056C (500 assay points***)	AL3056F (5000 assay points***)
AlphaLISA Anti-hFN Acceptor beads stored in PBS, 0.05% Proclin-300, pH 7.2	20 µL @ 5 mg/mL (1 brown tube, <u>white</u> cap)	50 µL @ 5 mg/mL (1 brown tube, <u>white</u> cap)	500 µL @ 5 mg/mL (1 brown tube, <u>white</u> cap)
Streptavidin (SA)-coated Donor beads stored in 25 mM HEPES, 100 mM NaCl, 0.05% Proclin-300, pH 7.4	80 µL @ 5 mg/mL (1 brown tube, <u>black</u> cap)	200 µL @ 5 mg/mL (1 brown tube, <u>black</u> cap)	2 x 1 mL @ 5 mg/mL (2 brown tubes, <u>black</u> caps)
Biotinylated Anti-hFN Antibody stored in PBS, 0.1% Tween-20, 0.05% NaN ₃ , pH 7.4	20 µL @ 500 nM (1 tube, <u>black</u> cap)	50 µL @ 500 nM (1 tube, <u>black</u> cap)	500 µL @ 500 nM (1 tube, <u>black</u> cap)
Lyophilized hFN*	3 µg (1 tube, <u>clear</u> cap)	3 µg (1 tube, <u>clear</u> cap)	3 µg (1 tube, <u>clear</u> cap)
AlphaLISA Immunoassay Buffer (10X)**	2 mL, 1 small bottle	10 mL, 1 medium bottle	100 mL, 1 large bottle

* Reconstitute lyophilized analyte in 100 µL Milli-Q® grade H₂O. The reconstituted analyte should be used within 60 minutes or aliquoted into screw-capped polypropylene vials and stored at -20°C for future experiments. Avoid freeze-thaw cycles. One vial contains an amount of analyte sufficient for performing 10 standard curves. Additional vials can be ordered separately (cat # AL3056S).

** Extra buffer can be ordered separately (cat # AL000C: 10 mL, cat # AL000F: 100 mL).

*** The number of assay points is based on an assay volume of 100 µL in 96-well plates or 50 µL in 96- or 384-well assay plates using the kit components at the recommended concentrations.

Sodium azide should **not** be added to the stock reagents. High concentrations of sodium azide (> 0.001 % final in the assay) might decrease the AlphaLISA signal. Note that sodium azide from the Biotinylated Antibody stock solution will not interfere with the AlphaLISA signal (0.0001% final in the assay).

Specific additional required reagents and materials:

The following materials are recommended:

Item	Suggested source	Catalog #
TopSeal™-A Plus Adhesive Sealing Film	PerkinElmer Inc.	6050185
EnVision®-Alpha Reader	PerkinElmer Inc.	-

Recommendations

- The volume indicated on each tube is guaranteed for single pipetting. Multiple pipetting of the reagents may reduce the theoretical amount left in the tube. To minimize loss when pipetting beads, it is preferable not to pre-wet the tip.
- Centrifuge all tubes (including lyophilized analyte) before use to improve recovery of content (2000g, 10-15 sec). Re-suspend all reagents by vortexing before use.
- Use Milli-Q® grade H₂O (18 MΩ·cm) to dilute 10X AlphaLISA Immunoassay Buffer and to reconstitute the lyophilized analyte.
- When diluting the standard or samples, change tips between each standard or sample dilution. When loading reagents in the assay microplate, change tips between each standard or sample addition and after each set of reagents.
- When reagents are added to the microplate, make sure the liquids are at the bottom of the well.
- Small volumes may be prone to evaporation. It is recommended to cover microplates with TopSeal-A Adhesive Sealing Films to reduce evaporation during incubation. Microplates can be read with the TopSeal-A Film.
- The AlphaLISA signal is detected with an EnVision Multilabel Reader equipped with the Alpha option using the AlphaScreen standard settings (e.g. Total Measurement Time: 550 ms, Laser 680 nm Excitation Time: 180 ms, Mirror: D640as, Emission Filter: M570w, Center Wavelength 570 nm, Bandwidth 100 nm, Transmittance 75%).
- AlphaLISA signal will vary with temperature and incubation time. For consistent results, identical incubation times and temperature should be used for each plate.
- The standard curves shown in this technical data sheet are provided for information only. A standard curve must be generated for each experiment.

Assay Procedure

IMPORTANT: PLEASE READ THE RECOMMENDATIONS BELOW BEFORE USE

- The protocol described below is an example for generating one standard curve in a 50 µL final assay volume (48 wells, triplicate determinations). The protocols also include testing samples in 452 wells. If a different amount of samples are tested, the volumes of all reagents have to be adjusted accordingly, as shown in the table below. These calculations do not include excess reagent to account for losses during transfer of solutions or dead volumes.
- The standard dilution protocol is provided for information only. As needed, the number of replicates or the range of concentrations covered can be modified.
- Use of four background points in triplicate (12 wells) is recommended when LDL/LLOQ is calculated. One background point in triplicate (3 wells) can be used when LDL/LLOQ is not calculated.

Format	# of data points	Final	Volume				Plate recommendation
			Sample	AlphaLISA Acceptor beads	Biotinylated Antibody	SA-Donor beads	
AL3056HV	100	100 µL	10 µL	20 µL	20 µL	50 µL	White OptiPlate-96 (cat # 6005290) White ½ AreaPlate-96 (cat # 6005560)
AL3056C	250	100 µL	10 µL	20 µL	20 µL	50 µL	White OptiPlate-96 (cat # 6005290) White ½ AreaPlate-96 (cat # 6005560)
	500	50 µL	5 µL	10 µL	10 µL	25 µL	White ½ AreaPlate-96 (cat # 6005560) White OptiPlate-384 (cat # 6007290) Light gray AlphaPlate™-384 (cat # 6005350)
	1 250	20 µL	2 µL	4 µL	4 µL	10 µL	Light gray AlphaPlate-384 (cat # 6005350) ProxiPlate™-384 Plus (cat # 6008280) White OptiPlate-384 (cat # 6007290)
	2 500	10 µL	1 µL	2 µL	2 µL	5 µL	Light gray AlphaPlate-1536 (cat # 6004350)
AL3056F	5 000	50 µL	5 µL	10 µL	10 µL	25 µL	White ½ AreaPlate-96 (cat # 6005560) White OptiPlate-384 (cat # 6007290) Light gray AlphaPlate-384 (cat # 6005350)
	12 500	20 µL	2 µL	4 µL	4 µL	10 µL	Light gray AlphaPlate-384 (cat # 6005350) ProxiPlate-384 Plus (cat # 6008280) White OptiPlate-384 (cat # 6007290)
	25 000	10 µL	1 µL	2 µL	2 µL	5 µL	Light gray AlphaPlate-1536 (cat # 6004350)

3 Step Protocol described below is for 500 assay points including one standard curve (48 wells) and samples (452 wells). If a different amount of samples are tested, the volumes of all reagents have to be adjusted accordingly.

1) Preparation of 1X AlphaLISA Immunoassay Buffer:

Add 5 mL of 10X AlphaLISA Immunoassay Buffer to 45 mL Milli-Q® grade H₂O.

2) Preparation of hFN analyte standard dilutions:

- a. Reconstitute lyophilized hFN (3 µg) in 100 µL Milli-Q® grade H₂O.
- b. Prepare standard dilutions as follows in 1X AlphaLISA Immunoassay Buffer (change tip between each standard dilution):

Tube	Vol. of hFN (µL)	Vol. of diluent (µL) *	[hFN] in standard curve	
			(g/mL in 5 µL)	(pg/mL in 5 µL)
A	10 µL of reconstituted hFN	90	3.0E-06	3 000 000
B	60 µL of tube A	120	1.0E-06	1 000 000
C	60 µL of tube B	140	3.0E-07	300 000
D	60 µL of tube C	120	1.0E-07	100 000
E	60 µL of tube D	140	3.0E-08	30 000
F	60 µL of tube E	120	1.0E-08	10 000
G	60 µL of tube F	140	3.0E-09	3 000
H	60 µL of tube G	120	1.0E-09	1 000
I	60 µL of tube H	140	3.0E-10	300
J	60 µL of tube I	120	1.0E-10	100
K	60 µL of tube J	140	3.0E-11	30
L	60 µL of tube K	120	1.0E-11	10
M ** (background)	0	100	0	0
N ** (background)	0	100	0	0
O ** (background)	0	100	0	0
P ** (background)	0	100	0	0

* Dilute standards in diluent (e.g. 1X AlphaLISA Immunoassay Buffer).

At low concentrations of analyte, a significant amount of analyte can bind to the vial. Therefore, load the analyte standard dilutions in the assay microplate within 60 minutes of preparation.

** Four background points in triplicate (12 wells) are used when LDL is calculated. If LDL does not need to be calculated, one background point in triplicate can be used (3 wells).

3) Preparation of 5X AlphaLISA Anti-hFN Acceptor beads (50 µg/mL):

- a. Prepare just before use.
- b. Add 50 µL of 5 mg/mL AlphaLISA Anti-hFN Acceptor to 4950 µL of 1X AlphaLISA Immunoassay Buffer.

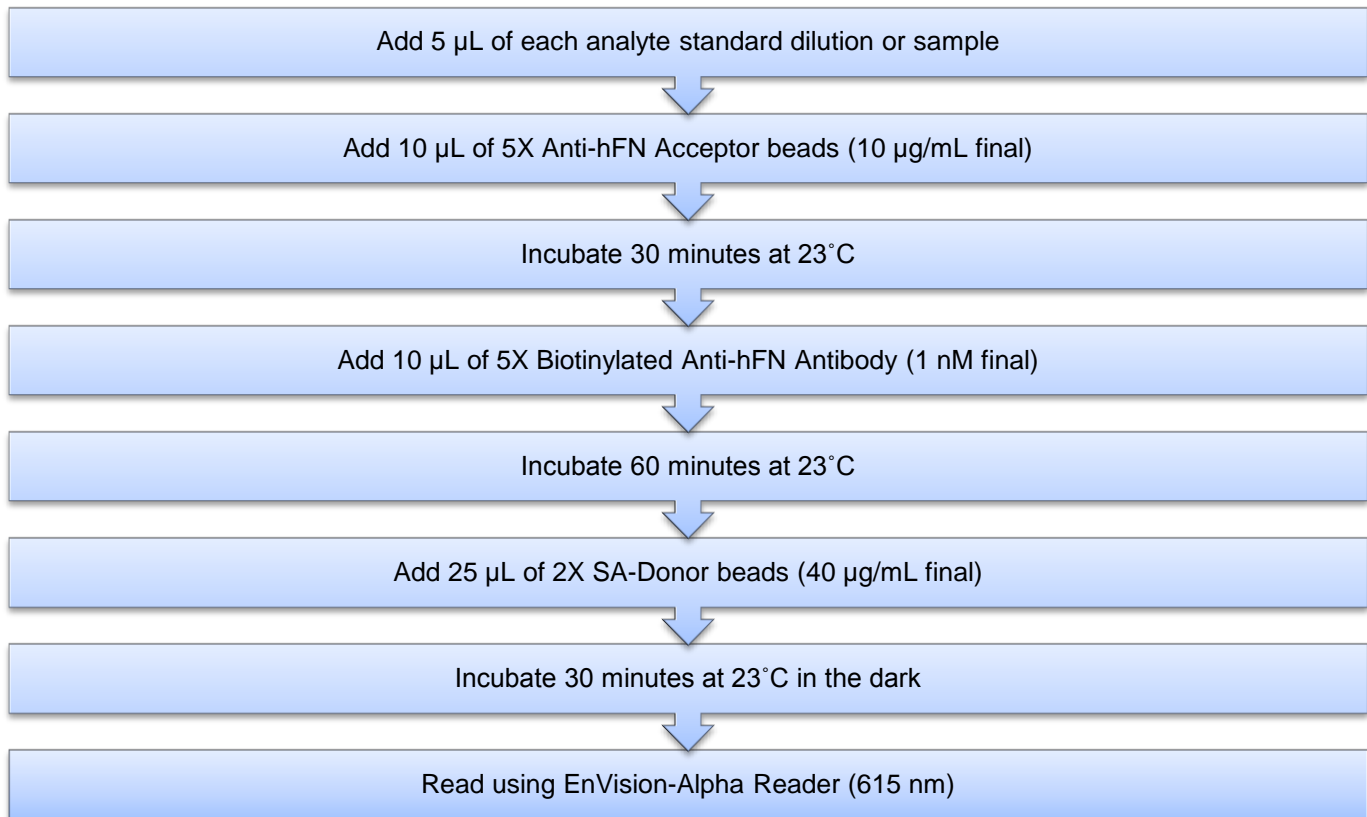
4) Preparation of 5X Biotinylated Anti-hFN Antibody (5 nM):

- a. Prepare just before use.
- b. Add 50 µL of 500 nM Biotinylated Anti-hFN Antibody to 4950 µL of 1X AlphaLISA Immunoassay Buffer.

5) Preparation of 2X Streptavidin (SA) Donor beads (80 µg/mL):

- a. Prepare just before use.
- b. Keep the beads under subdued laboratory lighting.
- c. Add 200 µL of 5 mg/mL SA-Donor beads to 12300 µL of 1X AlphaLISA Immunoassay Buffer.

6) In a white Optiplate (384 wells):



Data Analysis

- Calculate the average count value for the background wells.
- Generate a standard curve by plotting the AlphaLISA counts versus the concentration of analyte. A log scale can be used for either or both axes. No additional data transformation is required.
- Analyze data according to a nonlinear regression using the 4-parameter logistic equation (sigmoidal dose-response curve with variable slope) and a $1/Y^2$ data weighting (the values at maximal concentrations of analyte after the hook point should be removed for correct analysis).
- The LDL is calculated by interpolating the average background counts (12 wells without analyte) + 3 x standard deviation value (average background counts + (3xSD)) on the standard curve.
- The LLOQ as measured here is calculated by interpolating the average background counts (12 wells without analyte) + 10 x standard deviation value (average background counts + (10xSD)) on the standard curve. Alternatively, the true LLOQ can be determined by spiking known concentrations of analyte in the matrix and measuring the percent recovery, and then determining the minimal amount of spiked analyte that can be quantified within a given limit (usually +/- 20% or 30% of the real concentration).
- Read from the standard curve the concentration of analyte contained in the samples.
- If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

Assay Performance Characteristics

AlphaLISA assay performance described below was determined using the 3 step protocol using AlphaLISA Immunoassay Buffer (IAB). The analytes (standards) were prepared in IAB, DMEM + 10% FBS, RPMI + 10% FBS, or 100% FBS and all other components were prepared in IAB.

- Assay Sensitivity:

The LDL was calculated as described above. The values correspond to the lowest concentration of analyte that can be detected in a volume of 5 μ L using the recommended assay conditions.

LDL (pg/mL)	Analyte Diluents*	# of experiments
242	IAB	6
321	DMEM + 10% FBS	6
2195	RPMI + 10% FBS**	6
483	100%FBS	6

* The standard was prepared in these diluents and all other components were diluted in IAB. Note that LDL can be decreased (i.e. sensitivity increased) by increasing the volume of analyte in the assay (e.g. use 10 μ L of analyte in a final assay volume of 50 μ L).

**Significant (~10 fold) reduction in LDL, LLOQ, Maximum and Minimum signals, and signal to background ratio in RPMI+10%FBS. If the amounts of FN in RPMI+10%FBS culture media are > 2 ng/mL, the standard curve made in RPMI+10%FBS can be used.

- Assay Precision:

The following assay precision data were calculated from the three independent assays using two different kit lots. In each lot, the analytes were prepared in IAB, DMEM + 10% FBS, RPMI + 10% or 100%FBS. All other components were prepared in IAB. Each assay consisted of one standard curve comprising 12 data points (each in triplicate) and 12 background wells (no analytes). The assays were performed in 384-well format.

- Intra-assay precision:

The intra-assay precision was determined using a total of 16 independent determinations in triplicate. Shown as CV%.

FN	IAB	DMEM+ 10% FBS	RPMI+ 10% FBS	100% FBS
CV (%)	4	5	7	6

- Inter-assay precision:

The inter-assay precision was determined using a total of 3 independent determinations with 9 measurements for 3 ng/mL sample. Shown as CV%.

FN	IAB	DMEM+ 10% FBS	RPMI+ 10% FBS	100% FBS
CV (%)	8	10	9	12

- Spike Recovery:

Three known concentrations of analyte were spiked into IAB, DMEM + 10% FBS, RPMI + 10% or 100% FBS. All samples, including non-spiked diluents were measured in the assay. Note that the standard curves were prepared in IAB, DMEM + 10% FBS, DMEM + 10% FBS or 100% FBS. All other assay components were diluted in IAB.

Spiked FN (ng/mL)	% Recovery			
	IAB	DMEM+ 10% FBS	RPMI+ 10% FBS	100%FBS
63	90	91	89	111
6	83	85	84	80
0.6	96	100	98	104

- Specificity:

Cross-reactivity of the hFN Kit was tested using the following proteins at 100 ng/mL in IAB. The percent detected in the assay are shown in the table below.

Protein	% Cross-reactivity
Human Collagen Type I	0.1
Human Collagen Type III	0.0
Human Vitronectin	0.9
Human Fibronectin Fragment III.1	0.9
Mouse Fibronectin	0.1
Rat Fibronectin	0.1
Bovine Fibronectin	0.3

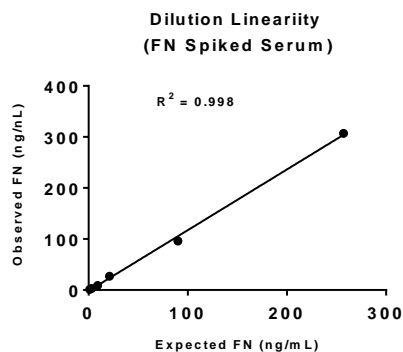
Human Serum Experiments

- Dilution Linearity

Neat Normal Human Serum and FN-spiked (300 ng/mL) Normal Human Serum samples were diluted with 100% FBS* and the assays were performed along with a standard curve prepared in FBS. The standard was prepared in FBS and all other reagents were prepared in IAB. Concentrations of FN in diluted samples were determined by interpolating to the standard curve. A large amount (262 ± 33 µg/mL) of FN was detected in normal human serum when the serum was

diluted ≥ 1000 fold. In the FN-spiked human serum samples, similar amount of FN ($292 \pm 16 \mu\text{g/mL}$) was detected and good dilution linearity ($R^2 = 0.998$) was obtained with the samples diluted ≥ 1000 fold. Accurate calculation of the spiked FN was impossible due to a large amount of endogenous FN ($>250 \text{ ng/mL}$) and small amounts of expected FN ($\leq 0.3 \text{ ng/mL}$) in the spiked serum that was diluted ≥ 1000 fold. Therefore, the expected (spiked) values in the table and graph below were replaced with the values of endogenous FN detected in normal serum samples at same dilutions.

Dilution Factor (DF)	FN Spiked samples (Expected, ng/mL)**	FN Spiked samples (Observed, ng/mL)
1000	257	307
3000	90	96
10000	21	27
30000	9	9
100000	3	3
300000	1	1



*The same assays were performed using the standard curve prepared in the IAB (data not shown). The results are identical, $287 \mu\text{g/mL}$ FN was detected in both neat serum and the spiked serum samples suggesting that IAB buffer can be used as diluent to prepare standard curve and to dilute serum samples.

- Spike Recovery

Three known amounts of FN were spiked into Normal Human Serum ($150, 450$ and $1500 \mu\text{g/mL}$ FN in spiked samples) and then the samples were diluted 30000-fold into 100% FBS. The standard was prepared in FBS and all other reagents were prepared in IAB. The spike recoveries of FN were determined and the results were shown in table below. *Recoveries were calculated after the no spike value was subtracted (in this case, $262 \mu\text{g/mL}$ human serum). Excellent recovery was achieved for all three samples.

Diluent: 100%FBS		
Spiked sample (Normal Human Serum)		
Spike ($\mu\text{g/mL}$)	Concentration ($\mu\text{g/mL}$)*	Recovery (%)
No spike	262	NA
150	135	90
450	426	94
1500	1458	97

Troubleshooting Guide

You will find detailed recommendations for common situations you might encounter with your AlphaLISA Assay kit at:

<http://www.perkinelmer.com/lab-products-and-services/application-support-knowledgebase/alphalisa-alphascreen-no-wash-assays/alpha-troubleshooting.html>

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